STRUCTURE AND STEREOCHEMISTRY OF MEXICANIN G, AN INTERMEDIATE IN THE BIOGENESIS OF HELENANOLIDES*

ALFONSO ROMO DE VIVAR, GUILLERMO DELGADO and EDUARDO HUERTA

Instituto de Química de la Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, Coyoacán 04510, México, D.F.

(Received 18 January 1985)

Key Word Index—Helenium mexicanum; Asteraceae; guaianolide; biogenesis; X-ray crystallography.

Abstract—The structure of mexicanin G, a guaianolide isolated from *Helenium mexicanum*, is established by means of an X-ray crystallographic analysis. Probably mexicanin G plays an important role in the biogenesis of helenanolides.

INTRODUCTION

In earlier articles [1, 2], the physical constants of the lactonic constituents of Helenium mexicanum H.B.K. were reported, and over the years this species has been the subject of several studies. The mexicanins A (1) [3], C (2) [4], D (3) [3, 5], E (4) [6, 7], H (5) [8] and I (6) [9] as well as helenalin (7)[10] were isolated from this plant and their structures were established by different analytical methods, but the structures of some mexicanins long remained undefined. Just recently, the structure of mexicanin F was determined as 8, which is a Diels-Alder adduct between the 11,13-double bond of mexicanin E (4) and the kinetic enol of the same nor-pseudoguaianolide [11]. All the mexicanins of authenticated structure possess a pseudoguaiane parent skeleton, a cyclopentanone, a C-12/C-8 y-lactone moiety (cis- or trans-) and belong to the helenanolide series (Me-C-10a). Here we report the structure and stereochemistry of mexicanin G (9), which lacked the general structural features of the mexicanins isolated from the same natural source since it is a guaianolide and therefore may be regarded as a key intermediate in the biogenesis of helenanolides.

RESULTS AND DISCUSSION

The molecular formula of mexicanin G (9) ($C_{17}H_{22}O_5$), mp 214-216°, $[\alpha]_D^{25} = 40^\circ$ (c 0.1), was determined by elemental analysis and mass spectrometry. The α -methylene- γ -lactone and acetate functionalities were established by their characteristic absorptions in the IR and ¹H NMR spectra (see Experimental). The two-proton complex signal centred at $\delta 4.78$ in the ¹H NMR spectrum indicates the presence of the secondary protons geminal to acetate and lactone groups. This spectrum also exhibits, in addition to the C-10 methyl doublet ($\delta 1.18$, 3H, d, J = 7 Hz), a low field singlet at $\delta 1.52$ (3H, s) of a methyl group, which indicates the presence of a C-3/C-4 tetrasubstituted epoxide ring in a guaiane skeleton, in agreement with the molecular formula. The low chemical shift of the

lactonic methine (δ 4.78) and biogenetic considerations suggest lactone closure to C-8. Irradiation experiments indicated that the acetate could not be attached to C-6. Of the remaining three possible positions for the acetate (C-2, C-3 and C-9), C-2 is favoured on biogenetic grounds, and therefore 9 is the most probable structure of mexicanin G, regardless of stereochemistry. Due to the uncertain configuration of C-1, C-2, C-3, C-4, C-8 and C-10, and the unavailability of material for chemical transformations, an X-ray analysis of mexicanin G was undertaken. Details of this analysis are given in the Experimental section and listings of pertinent crystallographic data are deposited at the Cambridge Crystallographic Data Centre. Figure 1 is a computer-generated drawing of the 1S,2S,4R,5S,7R,8R,10R-enantiomer and probably also represents the absolute stereochemistry on the basis that H-7 in sesquiterpene lactones from higher plants is α [12]. As can be seen from the drawings, the ring junctions of the guaianolide nucleus are cis at C-1-C-5 and at C-8-C-7, the acetate is at C-2 and α , the C-3/C-4 epoxide is α as well as the C-10 methyl group.

The co-occurrence of the guaianolide mexicanin G (9) and the pseudoguaianolides mexicanins A (1), C (2), E (4), H (5), I (6), F (8) and helenalin (7) is in agreement with the generally accepted biogenetic scheme of helenanolides which states that these compounds are formed from a trans-guaianolide cation A of Scheme 1, (which results from an intramolecular cyclization of a 4(5)-epoxygermacrolide [13] or a 4(5)-epoxy-melampolide [14, 15]). A would give the helenanolide skeleton B via a series of 1,2-migrations (due to antiperiplanar orientation of migrating groups). Mexicanin G (9) could be considered an intermediate of this multistep sequential rearrangement, since the eventual carbocation at C-5 C (formed via two consecutive 1,2-H shifts, from C-1 to C-10 and from C-5 to C-1, see arrows, Scheme 1) could stabilize itself by C-4/C-5 epoxide formation to give 9 (R = Ac). Alternatively, 9 could undergo an epoxide rearrangement and 1,2methyl shift to a helenanolide [16]. In addition, mexicanin G may be the biogenetic precursor of the sesquiterpene lactones similar to 2\alpha-tigloyloxy-dougaldiolide (10), isolated from the closely related genus Dougaldia, via the 11β , 13-epoxy-derivative of 9, as previously suggested [17].

2977

^{*}Contribution No. 738 Instituto de Química, U.N.A.M.

Fig. 1.

EXPERIMENTAL

Slow crystallization from EtOAc-hexane of the extensively purified sample of mexicanin G remaining from the earlier work [2] gave ca 5 mg of single crystals which melted somewhat higher than reported previously (mp 214–216°); IR $v_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$: 3039, 2964, 2929, 1755, 1725, 1659, 1602, 1453, 1383, 1237, 1088, 992; 1 H NMR (CDCl₃, 80 MHz): δ 6.28 (d, J = 3.1 Hz, H-13a), 5.60 (d, J = 3.0 Hz, H-13b), 4.98–4.58 (complex, 2H, H-8 and H-1), 3.50 (complex, H-7), 2.05 (3H, s, AcO), 1.52 (3H, s, 15-Me), 1.18 (3H, d, J = 7 Hz, 14-Me); EIMS 70 eV, m/z (rel. int.): 306 [M] $^+$ (< 1), 291 (3), 263 (10), 246 (12), 155 (9), 154 (11), 152 (10), 151 (12), 60 (83), 43 (100). Found: C, 66.80; H, 7.17; O, 26.26%. $C_{17}H_{22}O_{5}$ requires: C, 66.65; H, 7.24; O, 26.11%.

The crystals of mexicanin G (9) were monoclinic, space group P2₁, with a = 9.3385 (3) A, b = 8.9208 (3) A, c = 9.4978 (3) A, $\beta = 92.305$ (3) A and z = 2 (C₁₇H₂₂O₅). The intensity data were measured on a Nicolet R3m diffractometer (monochromated CuK_a radiation, ω scans, pulse-high discrimination). The size of

Scheme 1.

the crystal used for data collection was approximately $0.28 \times 0.38 \times 0.40$ mm. A total of 1104 accessible reflections were measured for $2\theta < 45$ of which 1018 were considered to be observed [I > 2σ (I)]. The structure was solved by direct methods [18] and was refined by blocked cascade matrix least square methods. In the final refinement, anisotropic thermal parameters were used for the non-hydrogen atoms and fixed isotropic temperature factor $U = 0.045 \text{ A}^2$ for the hydrogen atoms. The hydrogen atoms were included in the structure factor calculations. The final discrepancy indices are R = 0.0345 and $R_{\omega} = 0.0378$ for the 1018 observed reflections.

Acknowledgement—We thank Ms. Sc. Rubén A. Toscano of the X-ray Department, Instituto de Química de la Universidad Nacional Autónoma de México, for technical assistance.

REFERENCES

- 1. Romo de Vivar, A. and Romo, J. (1959) Chem. Ind. 882.
- 2. Romo de Vivar, A. and Romo, J. (1961) Ciencia (Méx) 21, 33.
- Herz, W., Romo de Vivar, A., Romo, J. and Viswanathan, N. (1963) J. Am. Chem. Soc. 85, 19.
- Herz, W., Romo de Vivar, A., Romo, J. and Viswanathan, N. (1963) Tetrahedron 19, 1359.
- 5. Romo, J., Romo de Vivar, A. and Aguilar, M. (1969) Bol. Inst.

- Quím. Univ. Nac. Autón. Méx. 21, 66.
- Romo, J., Romo de Vivar, A. and Herz, W. (1963) Tetrahedron 19, 2317.
- Ul-Haque, M. and Caughlan, C. N. (1967) J. Chem. Soc. B 355.
- Romo, J., Romo de Vivar, A. and Joseph-Nathan, P. (1966) Tetrahedron Letters 1029.
- 9. Domínguez, E. and Romo, J. (1963) Tetrahedron 19, 1415.
- 10. Herz, W. (1962) J. Org. Chem. 27, 4043.
- Romo de Vivar, A. and Delgado, G. (1985) Tetrahedron Letters 579.
- Fischer, N. H., Olivier, E. J. and Fischer, H. D. (1979) Fortschr. Chem. Org. Naturst. 38, 47.
- 13. Fischer, N. H. (1978) Rev. Latinoam. Quím. 9, 41.
- 14. Herz, W. (1977) Isr. J. Chem. 16, 32.
- Herz, W., Murari, R. and Blount, J. F. (1979) J. Org. Chem. 44, 1873.
- Herz, W., Govindan, S. V., Bierner, M. W. and Blount, J. F. (1980) J. Org. Chem. 45, 493.
- Bohlmann, F., Misra, L. N., Jakupovic, J., Robinson, H. and King, R. M. (1984) J. Nat. Prod. 47, 658.
- Sheldrick, G. M. (1981) SHELXTL Revision 3. An integrated system for solving, refining and displaying structures from diffraction data. University of Göttingen, Federal Republic of Germany.